

## Directions for Use, Collagen (2-D and 3-D)

### <u>Preparation of Collagen Gel for Cell Culture</u> (for 2-dimensional and 3-dimensional cell growth)

#### A. Materials

- 1. Collagen G (0.4% collagen solution) or Collagen G1 (0.5% collagen solution), stored at +2° to +10°C, MATRIX BioScience GmbH
- 2. 1M sodium hydroxide (NaOH), sterile, SIGMA-ALDRICH, S 2770-100 mL
- 3. 1M HEPES buffer, sterile
- 4. 10x concentrate of RPMI 1640 medium, sterile, SIGMA-ALDRICH
- 5. Sterile 50 mL beaker
- 6. LAF-unit, sterile air supply
- 7. Sterile pipettes
- 8. Sterile Petri dish, approx. 50 55 mm (approx. 2 ")
- 9. Incubator A (without CO<sub>2</sub>), Incubator B (with CO<sub>2</sub>)
- 10. Miscellaneous: pH meter, pH probe, sterile 0.1M HCl and/or sterile 0.1M NaOH
- 11. Cell suspension (high density) in growth medium

All solutions should be used refrigerated at +2°C to +10°C.

#### B. Preparation of neutralized collagen solution (aseptic, with sterile tubes under LAF-unit)

- 1. Mix 0.8 mL of 1M NaOH with 1.0 mL of 1M HEPES buffer (= 1.8 mL of solution A)
- 2. Mix 2 mL of 10x RPMI 1640 with 1.8 mL of solution A (= 3.8 mL of solution B)
- Mix 16 mL of 0.4-0.5 % collagen solution with 3.8 mL of solution B. Mix gentle and thorough, avoid trapping of air bubbles. The pH of the solution should ideally be at pH 7.8 - 8.0. If necessary adjust the pH by the addition of a few drops of sterile 0.1M HCl or 0.1M NaOH.
- Pipette (or pour) the neutralized collagen solution into a sterile Petri dish.
  Example: Pipette approx. 7 mL of the neutralized collagen solution into a sterile Petri dish with a diameter of 50-55 mm (approx. 2 inches) to completely cover the bottom of the Petri dish to a depth of 2-3 mm.

#### C. Gelation / fibrillogenesis by incubation of neutralized collagen solution

1. Incubate the neutralized collagen solution for a minimum of approx. 60 minutes or up to 16 hours (overnight) at 37°C to initiate and to complete collagen fibrillogenesis / gelation.

NOTE: Gelation occurs more rapidly in the absence of CO<sub>2</sub>. NOTE: Before use, collagen fibrils may alternatively be dried in the following manner:

- a) After gelation leave dish uncovered in a stream of sterile air under the LAF-unit overnight or until dry.
- b) Rinse fibrillar collagen film with sterile water in order to remove salts and to rehydrate the film.
- c) Collagen film can be used immediately for cell culture or allowed to dry again with sterile air and be stored for future use.



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### D. Types of cell preparations:

- **2-dimensional :** Prepare a neutralized collagen gel according to B.1. to B.4. and C.1. Disperse cells with medium on the surface of the collagen gel after gelation and incubate.
- sandwiched : Prepare a neutralized collagen gel according to B.1. to B.4. and C.1. Disperse cells with a small amount of medium on collagen gel after gelation (on the surface of the "bottom gel"). Pour a new layer of neutralized collagen solution (see B.1. to B.3) gently on top of the cell layer (for "top layer gel" formation). Incubate the neutralized collagen solution for a minimum of approx. 60 minutes or up to 16 hours (overnight) at 37°C to initiate and to complete collagen fibrillogenesis / gelation of the "top collagen layer". Continue incubation for cell growth of sandwiched cell layer.
- 3-dimensional : Prepare a neutralized collagen solution according to B.1. to B.3. Prior to gelation, suspend cells in neutralized collagen solution by mixing 10 % of a cell suspension in medium with 90% of the neutralized collagen solution. Incubate the mix of neutralized collagen solution and cells for a minimum of approx. 60 minutes or up to 16 hours (overnight) at 37°C to initiate and to complete collagen fibrillogenesis / gelation.

Continue incubation for cell growth.